African Swine fever in Poland

Grzegorz Woźniakowski, Zygmunt Pejsak, <u>Maciej Frant</u>, Krzysztof Niemczuk NATIONAL VETERINARY RESEARCH INSTITUTE IN PULAWY, POLAND

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National Reference ASF laboratory











Diagnostic capabilities



800-1000 samples/day (3 – shift labour day)







ASF diagnostic methods applied by NVRI

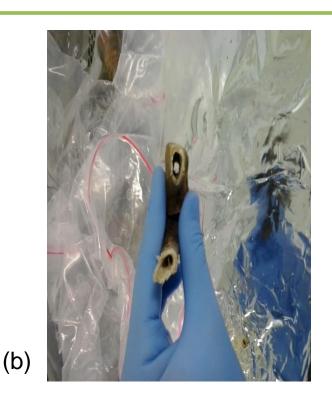
- Serological (ELISA, IB and IPT confirmatory assays,
- Molecular assays (real-time PCR),

ELISA, real-time PCR and IPT techniques are approved and accredited by Polish Accreditation Centre



Poor conditions of material sent for ASF diagnostic study





a) spilled and mixed blood samples, (b) a bone without any trace of bone marrow

Laborious and time consuming !!



ASF in wild boars



Passive surveillance - the most informative

	Part II + III (ASF- affected)							
Year	Fo	und Dea	d	Car accident				
	Tested	+	%	Tested	+	%		
2014	115	46	40%	68	0	0		
2015	130	67	51%	53	0	0		
2016	149	63	43%	95	3	3.15		
2017	471	282	60%	47	2	4.25		
(until 31.08)								



Active surveillance

Year	Part II + III (ASF - affected)				
lear	Tested	+	Prevalence		
2015	3387	14	0.41%		
2016	4221	24	0.56%		
2017 (until 31.08)	3528	66	1.87%		



Confirmation of 100% of results obtained by NRL at PIWet-PIB by EURL (Spain)

CONCLUSION*

- The presence of ASF has been confirmed throughout antibody and/or ASFV genome detection in all wild boar received from the cases occurred in Poland from December 2016 (case 165) up to March 2017 (case 250).
- The presence of ASF has been confirmed throughout antibody and/or ASFV genome detection in all wild boar received from the cases 107 to 162 which were occurred in Poland from August 2016 (case 107) up to December 2016 (case 162).

In Valdeolmos. Madrid (Spain), 24th May 2017

Dr. Carmina Gallardo Frontaura

Researcher, Laboratory Coordinator EU Reference Laboratory for ASF INIA-CISA Dr. Marisa Arias Neira

Technical Director
EU Reference Laboratory for ASF
INIA-CISA

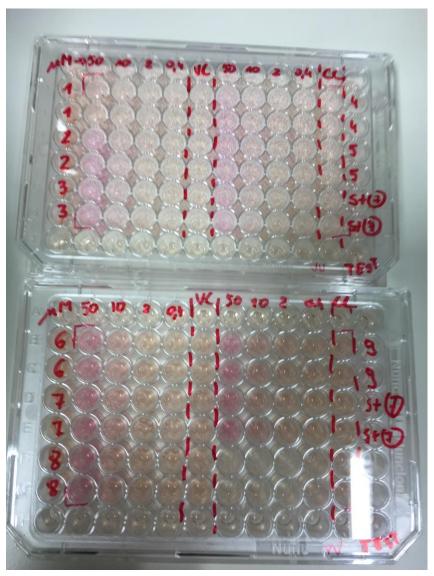
Approval Dr. Marisa Arias Neira

Technical Director INIA-CISA

Latest laboratory scientific activity



Examination of synthetic molecules inhibiting ASFV replication *in vitro*

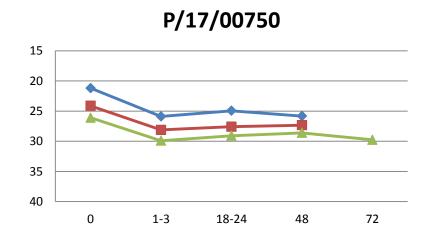


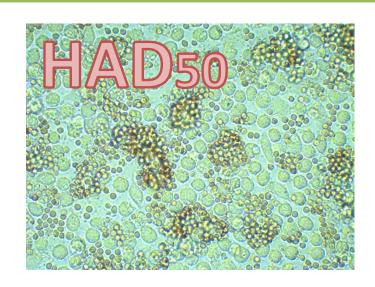
+ virus present - virus absent in 5 dpi duplicates

	50ι	иM	10	uМ	2uM		0,4uM	
VV03-								
0001	-	-	+	+	+	+	+	+
VV03-								
0002	-	-	-	-	+	+	+	+
VV03-								
0003	-	-	-	-	-	-	+	+
						+/-		
VV03-					+	(very		
0004	-	-	-	-	(weak)	weak)	+	+
			+	+				
VV03-			(wea	(wea				
0005	-	-	k)	k)	+	+	+	+
VV03-								
0006	-	-	+	+	+	+	+	+
VV03-					+			
0007	-	-	-	-	(weak)	+(weak)	+	+
					+/-	+/-		
VV03-					(very	(very		
8000	-	-	-	-	weak)	weak)	+	+
VV03-								
0009	-	-	-	-	-	-	+	+

Examination of ASFV infectivity/survivability after incubation of infected organs from pigs and wild boars in soil, liter and water (different conditions)



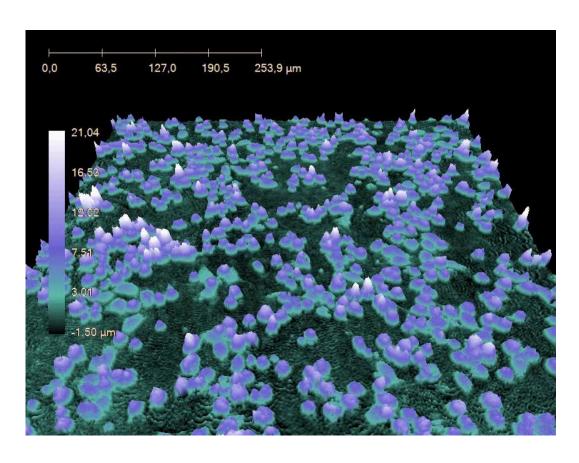






Construction of recombinant ASFV strain lacking genes involved in evasion host-immune response using CRISPR/Cas9





Observation of virus replication in infected pig macrophages using Holographic Microscope (no virus staining required)

Scientific papers published by the research team of the National Reference ASF Laboratory at the NVRI

EMERGING INFECTIOUS DISEASES®

African Swine Fever Epidemic, Poland, 2014-2015

Krzysztof Śmietanka, Grzegorz Woźniakowski, Edyta Kozak, Krzysztof Niemczuk, Magdalena Fraczyk, Łukasz Bocian, Andrzei Kowalczyk, Zygmunt Peisak

In Poland, African swine fever (ASF) emerged in February 2014; by August 2015, the virus had been detected in >130 wild boar and in pigs in 3 backyard holdings. We evaluated ASF spread in Poland during these 18 months. Phyloge-

very near (<1 km) the border with Belarus (6). As of Au 31, 2015, a total of 76 cases in wild boar and 3 outbre among domestic pigs had been found in 3 counties (b administrative regions of Poland).





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ORIGINAL ARTICLE

Development of cross-priming amplification for direct detection of the African Swine Fever Virus, in pig and wild boar blood and sera samples

M. Fraczyk¹, G. Woźniakowski¹, A. Kowalczyk¹, K. Niemczuk² and Z. Pejsak¹

- 1 Department of Swine Diseases, National Veterinary Research Institute (NVRI), Pulawy, Poland
- 2 National Veterinary Research Institute (NVRI), Puławy, Poland





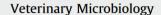
OPEN Polymerase cross-linking spiral reaction (PCLSR) for detection of African swine fever virus (ASFV) in

Received: 30 June 2016 Accepted: 03 January 2017 Published: 15 February 2017 pigs and wild boars Grzegorz Woźniakowski¹, Magdalena Frączyk¹, Andrzej Kowalczyk¹, Małgorzata Pomorska-Mól¹,

Krzysztof Niemczuk² & Zygmunt Pejsak¹

Veterinary Microbiology 193 (2016) 133-144

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journal homepage: www.elsevier.com/locate/vetmic



Evolution of African swine fever virus genes related to evasion of host immune response



Magdalena Fraczyka, Grzegorz Woźniakowskia,*, Andrzej Kowalczyka, Łukasz Bocianb, Edyta Kozak^a, Krzysztof Niemczuk^c, Zygmunt Pejsak^a

a Department of Swine Diseases, National Veterinary Research Institute, Partyzantów 57 Avenue, 24-100 Puławy, Poland

Department of Epidemiology and Risk Assessment, National Veterinary Research Institute, Partyzantów 57 Avenue, 24-100 Puławy, Poland

^c Chief executive, National Veterinary Research Institute, Partyzantów 57 Avenue, 24-100 Puławy, Poland **DE GRUYTER** OPEN

J Vet Res 60, 119-125, 2016 DOI:10.1515/jvetres-2016-0017

REVIEW ARTICLE

Selected aspects related to epidemiology, pathogenesis, immunity, and control of African swine fever

Grzegorz Woźniakowski, Magdalena Frączyk, Krzysztof Niemczuk, Zygmunt Pejsak

Department of Swine Diseases National Veterinary Research Institute, 24-100 Pulawy, Poland

Arch Virol (2016) 161:189-195 DOI 10.1007/s00705-015-2650-5

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BRIEF REPORT

Current status of African swine fever virus in a population of wild boar in eastern Poland (2014-2015)

Grzegorz Woźniakowski¹ · Edyta Kozak¹ · Andrzej Kowalczyk¹ · Magdalena Łyjak¹ · Małgorzata Pomorska-Mól¹ · Krzysztof Niemczuk² · Zygmunt Pejsak¹

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